

Molecular Genetic Technology Defines the Receptor Subtypes Responsible for Anesthetic and Cardiovascular Responses to alpha-2 Adrenergic Agonists

Toshiki Mizobe*, Takahiko Kamibayashi**, Guo Tianzhi**, Katsumi Harasawa**, Mervyn Maze**

Abstract

The analgesic effects of α_2 adrenergic agonists are mediated by activation of α_2 adrenoceptor in the locus coeruleus (LC) and spinal cord (SC). It is not known which of the three α_2 adrenoceptor subtypes is responsible for the analgesic effects. Using a genetargeting strategy, it is possible to determine their role in the analgesic action of dexmedetomidine, an α_2 agonist. The guide cannulae were sited stereotactically in the LC or intrathecally in male Sprague-Dawley rats. The antinociceptive effect of dexmedetomidine was measured using the tail flick latency (TFL) response. Separate cohorts of rats were administered the ODNs directed against either the α_{2A} or α_{2C} adrenoceptor subtypes which are present in the CNS. Control rats were treated with scrambled ODNs or saline. The TFL response to dexmedetomidine was measured, before, immediately after, and 8 days after the ODN administration. Immediately following α_{2A} ODN treatment, the TFL response to dexmedetomidine, either LC or SC, was significantly attenuated; this recovered to the pretreatment value after 8 days. α_{2C} ODN treatment did not change the analgesic response. The analgesic response to morphine was unaffected by

Key words: Adrenoceptor, α_2 adrenergic agonists, Analgesia, Dexmedetomidine

Introduction

 α_2 adrenergic agonists are efficacious, cardiovascular, anesthetic, and analgesic agents. However, the α_2 agonists do produce a variety of other pharmacologic actions affecting almost every organ system, especially the cardiovascular system causing quite troubling side-effects including hypertension, hypotension, and bradycardia¹). This plethora of α_2 adrenoceptormediated responses is due both to the ubiquity of this receptor class as well as to the existence of different receptor subtypes^{2,3}).

More than twenty years ago, Langer⁴) proposed that alpha adrenergic agonists such as clonidine were acting at a distinct presynaptic α_2 adrenoceptor to produce its sympatholytic effect. More recently it has become apparent that α_2 adrenoceptors can be located both pre and post-synaptically and that there are three distinct α_2 adrenoceptor subtypes, initially demonstrated by pharmacologic means and subsequently confirmed by molecular genetic techniques⁵).

Molecular genetic cloning studies in humans, rats and mice have shown that genes encode three distinct α_2 adrenergic receptor subtypes (Table 1). While phar-

 $[\]alpha_{2A}$ ODN treatment. These data strongly suggest that the α_{2A} adrenoceptor subtype mediates the analgesic responses to dexmedetomidine both in the LC and in the SC.

^{*}Department of Anesthesiology, Kyoto Prefectural University of Medicine, Kyoto, Japan

^{**}Departments of Anesthesia, VAPAHCS and Stanford University School of Medicine, USA

Table 1 Classification of α_2 adrenoceptors

pharmacologic nomenclature		α 2В	α2C
human genetic nomenclature	\alpha 2C10	α 2C2	Q 2C4
rat genetic nomenclature	RG20	RNG	RG10
mouse genetic nomenclature	MHC10	MHC2	MHC4
0			

macological studies have defined four subtypes named α_{2A} , α_{2B} , α_{2C} , and α_{2D} , α_{2D} represents a species homologue of the human α_{2A}^{6} (In this manuscript the term α_{2A} will be used to designate the $\alpha_{2A/D}$ subtype). While each of the three α_{2} adrenergic receptor subtypes are distributed in the central nervous syste $m^{7\sim15}$), the α_{2A} and α_{2C} subtypes predominate while the α_{2B} subtype is sparsely represented in the CNS and only in the diencephalon¹⁶).

The therapeutic index of α_2 agonists may be enhanced by developing strategies which avoid the unwanted actions while sustaining the desirable properties. If the salubrious properties are mediated by a different receptor subtype from that which mediates the undesirable effects, synthesis of subtype-selective α_2 agonists may represent a strategy for enhancing the therapeutic index of α_2 agonists. Pharmacologic¹⁷ and molecular genetic¹⁸ schemes represent two approaches that can be used to assign, unequivocally, an α_2 action to a particular subtype. Unfortunately, there are no truly selective pharmacologic ligands, neither agonists nor antagonists, which can be used to probe the individual receptor subtypes.

Recently, we and others have reported on the assignment of receptor subtype to individual responses using gene-targeting strategies. Kobilka's group has reported on cardiovascular studies of mice in which either the α_{2B} or α_{2C} adrenoceptor had been "knocked out." The α_{2B} knockout mice did not exhibit the acute hypertensive response to a bolus dose of dexmedetomidine. No cardiovascular phenotype was observed in the α_{2C} knockout mice. Using transgenic mice with dysfunctional α_{2A} adrenoceptors, Limbird's group was able to demonstrate that the α_{2A} adrenoceptor mediates the bradycardic and hypotensive effects²⁰).

We have addressed the anesthetic properties of the

 α_2 agonist, dexmedetomidine, and reported that the α_{2A} adrenoceptor subtype mediated the hypnotic response in the locus coeruleus of the rat¹⁸). Subsequently, using transgenic mice we confirmed that the α_{2A} adrenoceptor subtype was the anesthetic site of action for this species too²¹).

The α_2 agonists exert their analgesic properties by activating α_2 adrenoceptors in the neuraxis both supraspinally²²⁾ and in the spinal cord²³⁾, also α_2 agonists can produce pain relief at sites outside of the central nervous system^{24,25)}. Unlike opioids, the α_2 agonists do not exhibit potentially dangerous side-effects such as physical dependence and respiratory depression²⁶⁾. Using a gene-targeting strategy we now report on the α_2 adrenoceptor subtype responsible for the analgesic properties of dexmedetomidine in the locus coeruleus and spinal cord of the rat.

Methods

Synthesis of oligonucleotides

The following phosphodiester oligodeoxynucleotides (ODNs) were synthesized on ABI 394 and ABI 380B DNA Synthesizer by use of phosphoramidite chemistry (PAN Facility, Stanford, CA).

rat α_{2A} ODNs, 5'-ATG, GGC, TCC, CTG, CAG, CCG, GAT-3';

rat α_{2A} scrambled ODNs, 5'-CGA, GTT, GCC, TCA, AGC, GGT, CGC-3';

rat α_{2C} ODNs, 5'-ATG, GCG, TCC, CCA, GCG, CT-3';

rat α_{2C} scrambled ODNs, 5'-GGC, CTC, ACT, GCG, ACG, TC-3'

These represent sequences in the region immediately following the initiation codon of rat α_{2A} , and α_{2C} adrenoceptor subtypes. "Scrambled" ODNs contained the same nucleotides, but rearranged, and were used as controls for a non-specific ODN effect. Oligodeo-xynucleotides (ODNs), possess unique sequences relative to the entire genome and confer a degree of specificity which is lacking in conventional pharmacologic probes²⁷⁾. Earlier we showed that ODNs directed against the α_{2A} subtype resulted in a significant decrease in receptor expression (as defined by radiolabeled ligand binding) for α_{2A} with no

decrease in expression for α_{2C}^{18}). "Scrambled" ODNs did not affect receptor expression. Similarly, ODNs directed against the α_{2C} subtype selectively decreased expression only of α_{2C} receptor subtype¹⁸).

Cannulation for drug injection

The experimental protocol was approved by the Animal Care and Use Committee at the Veterans Affairs Palo Alto Health Care System. Male Sprague-Dawley rats, originating from the same litter, weighing 250-350g were used. The rats were stratified to match the distribution of the weights in the groups as closely as possible. All tests were performed between 10 a.m. and 4 p.m. The number of animals for each experiment is listed in the legends.

Locus coeruleus (LC) and intrathecal cannulations were performed as described before²²⁾. Briefly, halothane-anesthetized rats were placed in a stereotactic apparatus and the left LC was penetrated with a 24-G stainless steel cannula using coordinates according to the atlas of Paxinos and Watson²⁸⁾. The cannula was fixed in position with methylmethacrylate resin, and the animal was allowed to recover for 3 days before the experiment. The correct placement of the cannula was established by demonstrating that dexmedetomidine, 3.5 μ g / 0.2 μ l, produced loss of righting reflex (LORR). Only rats in which the previous administration of dexmedetomidine through the cannula resulted in LORR were used for subsequent studies.

For intrathecal cannulation, animals were anesthetized with halothane, an incision was made over the cervical spine, and a small puncture made in the dura mater. Polyethylene tubing (0.28 mm internal diameter) was threaded into the intrathecal space, 8.5 cm, so that the tip of the catheter was positioned at the lumbar enlargement. The tubing was sutured in place, and the skin sutured together over the tubing. The animal was allowed to recover for 3 days before the definitive experiment was initiated.

ODN administration

LC-cannulated rats received 3 injections on alternate days (day 1, 3, 5), with 5 nmol/0.2 μ l of phosphodiester ODNs, its "scrambled" control ODNs or 0.2 μ l

saline. Intrathecally-cannulated rats received 3 injections, on alternate days (day 1, 3, 5), with 5 nmol/10 μ l of phosphodiester ODN, its "scrambled" control ODNs or 10 μ l saline. These ODN treatment and recovery periods were selected based on the known half-life of the α_2 adrenergic receptor in the cerebral cortex²⁹.

Analgesia Testing

Analgesic response was measured by the tail-flick latency response. A high intensity light was focused on the rat's tail and the time for the rat to move its tail out of the light beam was automatically recorded (Tailflick apparatus, Columbus Instruments, Columbus, OH) and referred to as tail-flick latency. A different patch of the tail was exposed to the light beam on each trial to minimize the risk of tissue damage. The animals were placed on the heating blanket to maintain the body and tail temperature during the experiment. A cut-off time of 10 sec was predetermined, at which time the trial was terminated if no response occurred. Each tail flick latency data point consisted of a mean of three trials on an individual animal. Data are expressed as maximum percent effect (MPE) according to the following formula:

$$\text{MPE}\left(\%\right) = \frac{\left(\text{Postdrug latency}\right) - \left(\text{basal latency}\right)}{\left(\text{Cut-off latency}\right) - \left(\text{basal latency}\right)} \times 100\%$$

The analgesic response to an ED₅₀ dose of dexmedetomidine was performed on three separate occassions, namely before administration of ODNs, immediately after the last administration of ODNs, and 8 days after the last administration of ODNs.

To establish that the ODNs were acting at the receptor level to attenuate the analgesic response to dexmedetomidine, the analgesic effect of the opiate narcotic, morphine, was tested in rats treated with α_{2A} ODNs.

Statistical analysis

The data are expressed as mean \pm SEM. The results are analyzed by two-way analysis of variance with repeated measures, followed by post-hoc Scheffe' test. P < 0.05 is considered statistically significant.

Results

Dexmedetomidine, 3.5 μ g/0.2 μ l, LC, produced a similar % MPE in each cohort before ODN treatment. Immediately after α 2A ODN treatment the % MPE was significantly attenuated (Figure 1); the responses in the other treatment groups were unchanged. Following an 8 day recovery period after α 2A ODN treatment, the analgesic response to dexmedetomidine had recovered to the pretreatment values. The analgesic response to dexmedetomidine, 3.5 μ g / 0.2 μ l, LC, did not change after α 2C ODN treatment (Figure 2).

Dexmedetomidine, 1 μ g/10 μ l, intrathecal, produced a similar %MPE in each cohort before ODN treatment. Immediately following α_{2A} ODN treatment the %MPE was significantly attenuated (Figure 3); the

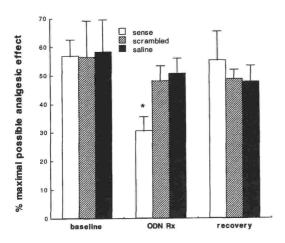


Figure 1 Effect of oligodeoxynucleotides (ODNs) directed against the α_{2A} adrenoceptor subtype on antinociceptive response to dexmedetomidine in the locus coeruleus.

Three cohorts of rat littermates, were stereotactically-cannulated, siting the tip of the needle in the locus coeruleus (LC). The antinociceptive response was represented by the percent of maximum possible prologation of the tail flick latency. The antinociceptive response to dexmedetomidine, 3.5 μ g, LC, was assessed before (baseline), immediately after (ODN Rx), and 8 days after (recovery) administering either α_{2A} ODN (n=5; 5 nmol/0.2 μ l), α_{2A} "scrambled ODNs" (n=5; 5 nmol/0.2 μ l), or saline (n=5; 0.2 μ l) three times, on days 1, 3, and 5. Data are expressed as mean \pm SEM. *p<0.05 when compared to "baseline" and "recovery" ODN treatment period.

other treatment groups were unchanged. Following an 8 day recovery period after α_{2A} ODN treatment, the analgesic response to dexmedetomidine had recovered to the pretreatment values. The analgesic response to dexmedetomidine, $1 \mu g / 10 \mu m$, intrathecal, did not change after α_{2C} ODN treatment (Figure 4).

The analgesic response to morphine, $0.65~\mu$ g/10 μ 1 intrathecally (determined to be the \approx ED₅₀ dose in pilot studies), which had produced a similar % MPE in both cohorts before ODN treatment, was unaffected by α _{2A} ODN treatment (Figure 5).

Discussion

 α_{2A} ODNs selectively, and reversibly, attenuated the analgesic response to dexmedetomidine whether administered supraspinally (Figure 1), or spinally (Figure 3). These data suggest that the α_{2A} adrenoceptor subtype is involved in the analgesic

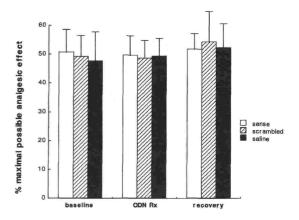


Figure 2 Effect oligodeoxynucleotides (ODNs) directed against the \$\alpha_{2C}\$ adrenoceptor subtype on antinociceptive response to dexmedetomidine LC.

Three cohorts of rat littermates, were stereotactically-cannulated, siting the tip of the needle in the locus coeruleus (LC). The antinociceptive response was represented by the percent of maximum possible prolongation of the tail flick latency. The antinociceptive response to dexmedetomidine, 3.5 μ g, LC, was assessed before (baseline), immediately after (ODN Rx), and 8 days after (recovery), administering either α_{2c} ODN (n=6; 5 nmol/0.2 μ I), α_{2c} "scrambled" ODN (n=5; 5 nmol/0.2 μ I), or saline (n=5; 0.2 μ I) three times, on days 1, 3, and 5. Data are expressed as mean \pm SEM.

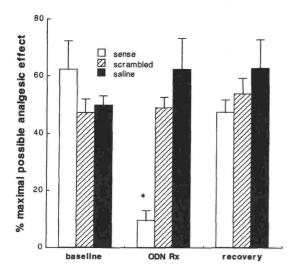


Figure 3 Effect of oligodeoxynucleotides (ODNs) directed against the α_{2A} adrenoceptor subtype on antinociceptive response to dexmedetomidine in the spinal cord (SC).

Three cohorts of rat littermates, were stereotactically-cannulated, siting the tip of the catheter at the lumbar enlargement of the intrathecal space. The antinociceptive response was represented by the percent of maximum possible prolongation of the tail flick latency. The antinociceptive response to dexmedetomidine, 1.0 μ g, SC, was assessed before (baseline), immediately after (ODN Rx) , and 8 days after (recovery), administering either α 2A ODN (n=5; 5 nmol/10 μ l), α 2A "scrambled" (n=4; 5 nmol/10 μ l), or saline (n=4; 10 μ l) three times, on days 1, 3, and 5. Data are expressed as mean \pm SEM. *p<0.01 when compared to "baseline" and "recovery" from ODN treatment period.

responses to α_2 agonists both in the LC and in the spinal cord. We were unable to prove that treatment with the α_{2A} ODNs reversibly changed the expression of α_{2A} adrenoceptor subtype in the LC and in the spinal cord since neither subtype-selective radiolabeled ligands, nor subtype-selective antibodies are available to perform the appropriate confirmatory test, namely, receptor subtype binding studies and in situ quantitative immunocytochemistry, respectively.

In lieu of definitive evidence for altered expression of α_{2A} adrenoceptors following α_{2A} ODN treatment, there exists considerable circumstantial evidence. Firstly, we earlier demonstrated that these same ODNs selectively decreased receptor expression in vitro¹⁸⁾.

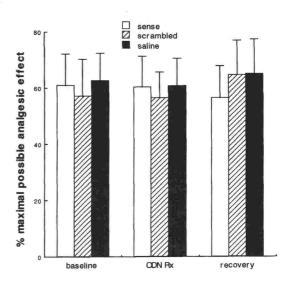


Figure 4 Effect of α_{2C} oligodeoxynucleotides (ODNs) on antinociceptive response to dexmedetomidine in the spinal cord (SC).

Three cohorts of rat littermates, were stereotactically-cannulated, siting the tip of the catheter at the lumbar enlargement of the intrathecal space. The antinociceptive response was represented by the percent of maximum possible prolongation of the tail flick latency. The antinociceptive response to dexmedetomidine, 1.0 μ g, SC, was assessed before (baseline), immediately after (ODN Rx) , and 8 days after (recovery), administering either α_{2C} ODN (n=5; 5 nmol/10 μ l), α_{2C} "scrambled ODN" (n=5; 5 nmol/10 μ l), or saline (n=5; 10 μ l) three times, on days 1, 3, and 5. Data are expressed as mean \pm SEM.

Secondly, only α_{2A} ODNs and not the "scrambled" ODNs diminished receptor expression in vitro and analgesic responses in vivo precluding the possibility that the behavioral changes are due to a non-specific ODN effect. Next, the behavioral effect are reversible over a time-course which is consistent with new receptor expression²⁹). Also, morphine and α_2 adrenergic agonists, which share the same signal transduction mechanism for analgesia in the spinal cord³⁰) are differentially affected by α_{2A} ODN treatment (Figures 1, 5). This final point suggests that the attenuated analgesic response to α_2 agonists is due to an alteration in the α_{2A} adrenoceptor itself and not due to a change in its post-receptor signal transduction

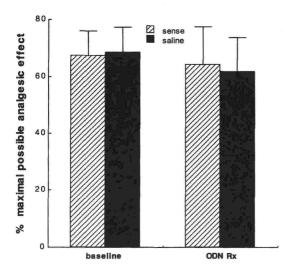


Figure 5 Effect of α_{2A} oligodeoxynucleotides (ODNs) on antinociceptive response to morphine in the spinal cord (SC).

Two cohorts of rat littermates, were stereotactically-cannulated, siting the tip of the catheter at the lumbar enlargement of the intrathecal space. The antinociceptive response was represented by the percent of maximum possible prolongation of the tail flick latency. The antinociceptive response to morphine, 0.65 μ g, SC, was assessed before (baseline), and immediately after (ODN Rx) administering either α_{2A} ODN (n=7; 5 nmol/10 μ l), or saline (n=6; 10 μ l) three times, on days 1, 3, and 5. Data are expressed as mean \pm SEM.

mechanism.

In studies using pharmacologic probes, Millan's group suggested that antinociceptive responses are mediated by α_{2A} adrenoceptors³¹⁾. Although their experimental paradigm differed with respect to species (mice vs rats) and nociceptive test (writhing and hot-plate vs tail-flick latency), we have corrobrated their conclusions. Initially, Wikberg's group suggested that only the α_{2A} adrenoceptor subtype was expressed in the spinal cord³²⁾, recently, Yaksh suggested that an additional α_2 adrenergic receptor is also capable of mediating the antinociceptive response to α_2 agonists³³⁾. Perl suggested that only α_{2A} adrenergic receptor subtype is expressed in dorsal root ganglion cells³⁴⁾. We can find no role for an antinociceptive response to dexmedetomidine via the α_{2C} adrenergic receptor subtype. This does not preclude the possibility that the antinociceptive effect of a different α_2 agonist measured by a different paradigm may act via a different receptor subtype.

The α_2 adrenergic agonists may be effective analgesics for both acute and chronic pain states. However, none of the clinically-available α_2 agonists can discriminate between the three α_2 adrenoceptor subtypes. This relative non-specificity may be the cause of troublesome side-effects such as hypertension. The acute hypertension that follows rapid bolus administration of α_2 agonists is probably due to activation of the α_{2B} adrenoceptor¹⁹, a subtype which is not found to any great extent in the neuraxis of rodents¹²⁾. If our data are extrapolatable to humans then the full potential of this drug class for analgesia may only be realized once ligands which have specificity for the receptor subtype(s) responsible for the salubrious effects thereby avoiding sites responsible of producing side-effects. Therefore, it is important to define, unequivocally, the receptor subtype(s) responsible for the anesthesia-related properties. This current study, taken together with the recent "knockout" and

"transgenic" studies, suggests that the α_{2A} adrenoceptor subtype could be the target for subsequent drug development of α_{2A} agonists for analgesic use.

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